

CENTRIFUGAL PARTITION CHROMATOGRAPHY (CPC) – A NOVEL METHOD OF SEPARATION AND PURIFICATION OF NATURAL PRODUCTS - A SHORT REVIEW

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Abstract

As technology has evolved, the field of liquid chromatography has developed, bringing new, alternative and high level methods of separation, isolation and purification of the natural products present in the natural plant extracts, replacing the traditional chromatographic methods. In this direction, until now, it has been observed that the Centrifugal Partition Chromatography (CPC) presents a high precision in isolating the individual compounds. Thus, we chose to study this topic in the present review. Currently, there is a growing interest in the use of CPC in the sustainable isolation and purification of cannabidiol (CBD) from Cannabis plants due to the extensive medical benefits of cannabis compounds (i.e. epilepsy, multiple sclerosis, etc.); atractylenolide I, selina4(14),7(11)-dien-8-one and (6E,12E)-tetradeca-6,12-diene-8,10-diyne-1,3-diol diacetate from *Atractylodis Rhizoma Alba*; xylindein from *Chlorociboria aeruginosa*; diphlorethohydroxycarmalol and octaphlorethol A from *Ishige okamurae*; Caulerpenyne from *Caulerpa taxifolia*, etc., most of them with antitumoral activities. Therefore, the challenge of the researchers is to obtain natural compounds through optimization protocols (including columns, sample properties in the solvent system, physical properties of the stationary and mobile phase, parameters of the instrument, etc.), saving time and in an economically cheaper manner. In conclusion, it can be stated that CPC is superior to the traditional methods, extracting compounds faster with high purity (99%), with a higher recovery rate (95%); also, it is environmentally sustainable, contributing efficiently to the pharmaceutical industry development.

Keywords: Centrifugal Partition Chromatography (CPC), natural products purification, separation.

1. INTRODUCTION

One of the most recently used and advanced techniques in liquid–liquid chromatography is centrifugal partition chromatography (CPC). It is a technique derived from counter-current chromatography, being also known as Hydrostatic Countercurrent Chromatography (HCC) (Foucault, 1995). Today, the purification of natural compounds present in the natural plant or fruits extracts is a complex process, which involves the usage of techniques and devices with high precision in separating and isolating the individual bioactive compounds. CPC is a novel technique which brings bright results in separation, isolation and purification of natural liquid extracts

mixtures, in a shorter time. This procedure is not using a silica column or support media, but two immiscible liquid phases (mobile and stationary phases) (Berthod et al., 2015). The partition between this two immiscible phases of a solvent system is the main principle of the CPC. The centrifugal field created keeps the stationary phase in the column, when the rotor starts to spin. The stationary phase is distributed and retained inside of the rotor and the mobile phase which contains the solutes to be extracted is pumped through under pressure as a mobile phase, crossing through stationary phase (Berthod et al., 2015). Based on the partition coefficients (K_D) and selectivity (α), the solutes present in the mobile phase inside the CPC (rotor) column are separated and isolated. The partition coefficient represents a ratio between the solutes concentration in the stationary and mobile phases. This may explain the fact that the mixture compounds with higher affinity for the mobile phase are eluting earlier than others with higher affinity for stationary phase.

Researchers are working on the optimization protocols, using all the advantages of the CPC compared with the traditional separation techniques, including: the low solvent consumption; performance, purity and recovery at high levels; a low running time at a high flow rate; the impossibility of denaturation and sample loss.

The aim of this review paper is to conduct a literature study using public libraries, which contain research articles from the last 5 years (2016 – present). This is a review of research on the optimization protocols and bioactive compounds types separated, isolated and purified from synthetic compounds mixtures and/or plant and fruits material by CPC. First of all, the brief CPC principles will be presented regarding the operation modes and optimisation steps, as well as the components. More, will be presented the biphasic solvent system types, best fitted, according to the researchers' works. Finally, a review of the recently conditions and examples of isolation, separation and purification of bioactive compounds by CPC from synthetic compounds mixtures and/or plant and fruits material os presented.

2. BRIEF CPC PRINCIPLES

2.1. Brief CPC device description

A CPC device has many advantages as: high recovery of the sample, various types of solvent systems, shorter running time, high injection volume and easy scale-up. A CPC system works as a preparative LC column, analogous to a preparative high performance liquid chromatography (HPLC) column, and its main components are the rotor (or column) and the rotary seals. Generally a CPC rotor is composed of individual disks, which contains twin cells linked by channels. Inside of the twin cells the liquid stationary phase is retained. Both sides of each twin cell have channels for the entrance and the exit of the mobile-phase.

2.2. Operation modes

The CPC operation modes are two, the ascending mode and the descending mode. In the ascending mode the lightest/upper phase is the mobile phase, and in the descending mode the heaviest/lower phase is the mobile phase. There is the possibility to change the flow direction during the process and to operate in dual-mode, ascending mode mixed with descending mode or vice-versa. In order to occur the separation the K_D is ranged between 0.5 and 5 (Marlot et al., 2017). If the K_D value is lower then 0.5 there is no separation and the analyte is retained in the mobile phase. In the opposite case, K_D value is higher then 5, the analyte is retained in the stationary phase. Therefore, an extremely important step in operating the CPC is the selection of solvent system.

2.3. Optimization steps in CPC

Optimization processes involve a number of mechanical, chemical and temporal factors. Most common factors are: rotational speed, flow rate, solvent system, elution step duration and extrusion step duration. These factors depend on the operation mode selected. The solvent systems can be optimized using the Arizona phase system family or trying different experimental conditions.

In order to optimize the purification process studies reported the usage of strong ion exchange CPC (IX-CPC) (Boudesocque et al., 2017).

The separation and isolation processes are influenced by parameters as sample properties (partition coefficients of the molecules, bio-active compounds concentration), solvent system (interfacial tension, densities, viscosities), instrument (shape, volume, cells material, size) and method (operation mode, flow rate, rotational, injection volume) (Bojczuk et al., 2017).

3. BIPHASIC SOLVENT SYSTEM

As mentioned before, biphasic solvent systems selection is an important step in the CPC separation. The most used solvents are heptane, ethyl acetate, methanol and water. There are several studies that have used biphasic solvent systems based on deep eutectic solvents for the separation and isolation of compounds from a mixture (Bezold et al., 2017). For a better solvent system selection, Bezold et all. proposed (Bezold et al., 2017) the predictive thermo-dynamic model COSMO-RS (Conductor-like Screening Model for Realistic Solvation).

The most common types of biphasic solvent systems for CPC contain a mixture of three or four solvents, as Arizona phase system composed of heptane (or hexane), ethyl acetate, methanol and water; the Oka scale composed of hexane, ethyl acetate, n-butanol, methanol, and water; and the acetone scale which is composed of heptane, toluene, acetone, and water (Boonloed et al., 2016). The biphasic solvent systems found in the research are listed in Table 1.

4. ISOLATION, SEPARATION AND PURIFICATION OF BIOACTIVE COMPOUNDS BY CPC FROM SYNTHETIC COMPOUNDS MIXTURES AND/OR PLANT AND FRUITS MATERIAL.

Isolation, separation, recovery and purification of bioactive compounds by CPC are important tools in nowadays researches. The bioactive-compounds isolated, recovered, purified or separated by CPC methods are presented in the Table 1.

During the last years, the CPC is intense applied in various fields, and a study conducted by Phansalkar (Phansalkar et al., 2018) reports the fact that CPC enabled selective enrichment of trimeric and tetrameric proanthocyanidins for biomaterial development.

Tabel 1. Isolation, separation and purification of bioactive compounds by CPC

| Biphasic solvent system | Ratio (v/v) | Isolation/ Separation/ Fractionation/ Purification | Compounds |
|---|-------------|--|--|
| n-Heptane/Ethanol/Choline chloride – levulinic acid | 1/1.33/1 | separation | α, γ – tocopherol from tocopherol mixture (Bezold et al., 2017). |
| n-Heptane/Ethanol/Choline chloride – 1,4-butanediol | 1/1.33/1 | | |
| Methyl tert-butyl | 2/1/2/5 | purification | dirucotide (69% recovery; |

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|--|------------------------|----------------------------|--|--|
| ether/Acetonitrile/ Butanol/Water | n- | | | 98% purity) from dirucotide 84.3% (Boudesocque et al., 2017). |
| Methanol/n-Hexane/Water | 5/5/0.8 | isolation, purification | | valtrate and 7-homovaltrate (97%) from <i>Centranthus ruber</i> L. Roots (Chami et al., 2018). |
| Methyl tert-butyl ether/Water | 1/1 | fractionation | | water soluble phase of a fast pyrolysis bio-oil (Dubuis et al., 2019). |
| Heptane/Ethyl acetate/Methanol/Water | 1/2/1/2 | separation | | fast pyrolysis oil produced from softwood sawdust (Le Masle et al., 2018). |
| n-Hexane/Ethyl acetate/ Ethanol/Water | 8/2/5/5 | isolation purification | | Acidic cannabinoids (>45% recovery; >95% purity) from <i>Cannabis sativa</i> L. (Popp et al., 2019). |
| n-Heptane/Ethyl acetate/ Methanol/Water | 4/1/4/1 (Arizona U) | purification | | Caulerpenyne from <i>Caulerpa taxifolia</i> (Sfecci et al., 2017). |
| n-Heptane/Ethyl acetate/ Methanol/Water | Arizona K-Q and T-Z | fractionation | | Enrichment of anthraquinones from <i>Kniphofia uvaria</i> seeds (Duval et al., 2016). |
| n-Heptane/Tetrahydrofuran/ Methyl ethyl ketone/Acetonitrile/Acid acetic/Water | 2/5/2/2/ 0.1/2 | isolation purification | | Xylindein from <i>Chlorociboria aeruginosa</i> (Boonloed et al., 2016). |
| Toluen/Acetonitrile/Water | 4/1/5 | purification | | Catharanthine and vindoline from aerial parts of <i>Catharanthus roseus</i> (Kotland et al., 2016). |
| Heptane/Methanol/Acetonitrile | 6/1/2 | separation purification | | Pyrethrins (99%) (Pyrethrin I, cinerin I, jasmolin I) from flowers of <i>Chrysanthemum cinerariaefolium</i> (Wong and Glinski, 2017). |
| Heptane/Terbutylmethyl ether/Acetonitrile/Water | 8/1/5/1.5 | separation purification | | Pyrethrins (99%) (Pyrethrin II, cinerin II, jasmolin II) from flowers of <i>Chrysanthemum cinerariaefolium</i> (Wong and Glinski, 2017). |
| n-Hexane/Ethyl acetate/Methanol/Water | 8/2/8/2 | isolation purification | | (6E,12E)-tetradeca-6,12-dien-8,10-diyne-1,3-diyI diacetate, atracylenolide I, selina- |

| | | | |
|--|-------------------|----------------------------|---|
| | | | 4(14),7(11)-dien-8-one from <i>Atractylodis Rhizoma Alba</i> (Kim et al., 2018). |
| Heptane/Methyl ether/Ethanol/Water | ter-butyl 4/1/4/1 | purification | Carnosol from <i>Rosmarinus officinalis</i> (Bouju et al., 2016) |
| Hexane/Ethyl acetate/Methanol/Water | 3/2/3/2 | purification | Carnosic acid ($96.1 \pm 1\%$) and carnosol ($94.4 \pm 0.9\%$) from <i>Rosmarinus officinalis</i> (Grace et al., 2017). |
| Toluene/Acetic acid/Water | 30/24/50 | separation purification | Aflatoxins produced by <i>Aspergillus parasiticus</i> and <i>Aspergillus flavus</i> (Endre et al., 2019). |
| Methyl ter-butyl ether/Water | 1/1 | isolation purification | Leontopodic acid A and 3,5-dicaffeoylquinic acid from Edelweiss plant extract (Marlot et al., 2017). |
| <i>n</i> -Hexane/Ethyl acetate/Ethanol/Water | 5/5/5 8/2/8/2 | separation isolation | 9-hydroxy isoegomaketone, isoegomaketone, perilla ketone from <i>Perilla frutescens</i> var. <i>crispa</i> leaves extract (Nam et al., 2019). |
| Hexane/Ethyl acetate/Methanol/Water | 1/9/1/9 | isolation | Proanthocyanidins from <i>avocado peels</i> (Torres et al., 2018). |

5. CONCLUSIONS

CPC represents a reproducible and robust technique for large-scale separation. The CPC method is developed for the preparative scale purification of bioactive compounds from mixtures of compounds. In the field of the drug development, natural products still contribute maintaining the interest of researchers for their isolation and purification.

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